

# The Role of Environmental Factors and Plant Growth Regulators on Grapes Coloration

### Raed S. Shehata<sup>1</sup>

<sup>1</sup> Department of Agriculture, Ministry of Agriculture and land reclamation, Damanhour, Egypt.

How to cite: Shehata, R.S. (2024). The Role of Environmental Factors and Plant Growth Regulators on Grapes Coloration. Viticulture Studies (VIS), 4(2): 9-20. <u>https://doi.org/10.52001/vis.2024.24.9.20</u>

**Article History:** Received: 08.03.2024 Accepted: 10.05.2024 First online: 06.06.2024

#### Corresponding Author raedsalem8882001@gmail.com

Keywords Grapes coloration Environmental factors Plant growth regulators Anthocyanin

### Introduction

### Abstract

Grapevine is the world's commercial horticulture crop. Poor coloration of grapes is one of the problems in grape production. Climate changes could negatively affect grape quality by reducing color formation. The effect of high temperatures on anthocyanin content and composition in the main red-producing grapes would help estimate their phenotypic change and perhaps predict whether they will be able to sustain the attributes of high-quality grapes in the context of climate change. Climate changes played an important role in lack sufficient berry color especially high temperature inhibited L-phenylalanineammonialyase (PAL). Lphenylalanineammonialyase improved accumulation of anthocyanin in the skin of grape berries. Environmental factors like light, temperature and irrigation are effective in either grapes coloration or discoloration. Plant growth regulators like Brassinosteroids (BRs), Jasmonic acid (JA), Salicylic acid (SA) and Abscisic acid (ABA) are accelerating the accumulation of anthocyanin in grapes berry skin. So it is very important to know how grapes coloration, the effect of plant growth regulators and environmental factors on grapes coloration.

An important perennial crop grown in many countries is grapevine mainly for wine production, but also for table grapes and raisins (Alston and Sambucci, 2019). The main components that determine the composition of grape berries are sugars, organic acids, and various secondary metabolites such as tannins, flavonols, anthocyanins, aroma precursors, and volatile compounds, which is significant for grape growers (Conde et al., 2007). The maturation process is influenced by both and internal factors such external as temperature, light, phytohormones, and plant water status (Gao-Takai et al., 2017; Sugiura et al., 2018; Azuma, 2018). Anthocyanins accumulate into the skin of grape berries due to a variety of environmental stimulation, such as developmental signals, environmental stresses

such as (light, irrigation, temperature, etc.), and plant growth regulators (Boss and Davies, 2009). Because well-pigmented grapes are often preferred by customers, skin color is a crucial characteristic that serves as the foundation for selection in breeding projects. Farmers appreciate these fruits' great marketability so, The skin color of berries is determined by the composition and quantity of anthocyanins. By altering the expression of genes involved in the anthocyanin biosynthesis pathways plant growth regulators can be used to improve the color of red grapes even at low concentrations of these compounds. From veraison until harvest anthocyanin biosynthesis and accumulation in skin cells occur primarily under genetic control (Costantini et al., 2015; Gagné et al., 2011).

This review observed that recent studies of plant

growth regulators and environmental factors on The regulation of anthocyanin production in grape berry skin to give growers a workable and practical regime for producing grape clusters with high quality.

# Enviromental Factors Affecting on Grapes Coloration:

### Light

According to Matus et al. (2009), exposure to light is a significant factor that affects grape coloration through the up-regulation of several genes including those involved in the biosynthesis of anthocyanins. The amount of light that reached the berries was correlated with the color of the clusters (Ma et al., 2019). Plant cells absorb light through phytochromes or photoreceptors which are then used by a number of metabolic pathways within the plant. PAR or photosynthetically active radiation is the 400–700 nm solar radiation range that plants can use for photosynthesis the process of turning light energy into usable energy. Each wavelength in the white light spectrum which spans from 380 nm to 780 nm, is received by phytochromes in plant cells and serves a distinct purpose (Demotes-Mainard et al., 2016). According to Cheng et al. (2015), grape berries' anthocyanin content is enhanced when they are exposed to blue light which is achieved by covering the greenhouse with blue plastic film. According to Cohen et al. (2012), berries heated to 20.5°C and cooled to ±8°C changed the initial rates of proanthocyanidin accumulation. On the other hand, the total proanthocyanidin accumulation appears to be more dependent on the development of berries during a specific season than it is on the thermal time. By modifying the expression of genes affected in the flavonoid biosynthesis pathway, cultivar, temperature, and light conditions can also assess the anthocyanin content in grape skin (Azuma et al., 2012). Light exclusion reduces the concentration and modifies the composition of grape anthocyanins by altering the expression of genes involved in the process of anthocyanin biosynthesis and transport in a cultivar and tissue specific manner (Guan et al., 2016). Numerous viticulture practices have been employed in previous studies on light exposure and cluster shading.

Some of the techniques utilized are the use of plastic netting (Chorti et al., 2010), shade cloths (Caravia et al., 2016; Greer and Weedon, 2013), and some leaf removal techniques surrounding the grape cluster (Lee and Skinkis, 2013; Chorti et al., 2010). (Guan et al., 2014) demonstrated that as anthocyanins accumulated during berry ripening the skin's ability to transmit light decreased. As a result, the berry flesh's shading effect is enhanced by the colored skin.

### Temperature

Climate change related high temperatures and decreased precipitation lead to advanced maturation which balances grape sugars and impacts phenolic maturity and grape composition (Poni et al., 2018). Farag et al, (2012) observed that, in comparison to non-dissipation accumulation heat, ethrel formulation at (400 ppm) in the presence of ethanol at (5%) at 15-20% berry coloration improved berry color and quality of "Crimson Seedless" grapes at harvest. According to Farag et al. (2011) in order to prevent accumulation of heat it is recommended to apply EDTA or ethanol two safe chemicals, under an open canopy in between rows of "crimson" seedless vines in order to increase the effectiveness of ethylene at 400 ppm. During ripening high temperatures (HT) typically suppress the anthocyanin accumulation that gives grape berries their color while low temperatures encourage it (Shinomiya et al., 2015). One of the major environmental factors regulating a cultivar's anthocyanin profile is temperature (De Rosas et al., 2017). Knowing the molecular mechanisms underlying temperature's effects on grape coloration during ripening is crucial because summertime HTs may reduce the color of wine and table grapes by inhibiting the biosynthesis of anthocyanins in grape berries (De Orduña, 2010; Barnuud et al., 2014; Fraga et 2012). Temperature is possibly the al., environmental factor that affects anthocyanin accumulation in grapes the most (Jones et al., 2012). According to a recent study by Gaiotti et al. (2018), A single season of the two seasons saw an increase in anthocyanin accumulation in grapes exposed to cool night temperatures (10-11°C, as compared with 15–20°C in the control) from 12 days before veraison until the end of veraison. Temperature regimes also had an impact on the composition of anthocyanins. High temperature treatments increased the proportion of acylated and methoxylated anthocyanins (De Rosas et al., 2017). According to recent research, flavonol accumulation was negatively impacted by high temperatures (30-40°C) during the berry ripening process (Degu et al., 2016; Pastore et al., 2017). According to Gaiotti et al. (2018) and industry anecdotal information grape flavonoid concentration is influenced by variations in day and nighttime temperatures with larger variations favoring flavonoid concentration. The fact that nighttime temperatures have increased more guickly than daytime temperatures over the past 50 years due to climate change makes this topic even more crucial today and points to future years of declining day-to-night temperature differences (Stocker et al., 2014). In particular, it was discovered that high temperatures in some cultivars altered the physiology and chemical composition of the berries as well as reducing their color (Sadras and Petrie, 2011). Furthermore, red wines made from the cultivars Barbera and Croatina were found to have lower resveratrol contents when fruit ripened at high temperatures (Rocchetti et al., 2021).

### Irrigation

It has been discovered that irrigation of vineyards impacts the biosynthesis of phenolics, which is a common practice in arid and semi-arid regions of the world (Cohen and Kennedy, 2010). When water stress or limitation occurs (i.e., before or after veraison). 'Crimson Seedless' coloration is being improved by post-veraison regulated deficit irrigation (RDI) techniques (Faci et al., 2014).

Grapevine productivity may be restricted by the distribution of rainfall and water availability, particularly towards the end of the growth cycle (Fraga et al., 2016, 2018). Different management practices like deficit irrigation (DI) can help to increase water use efficiency (WUE) maintain or improve wine quality and stabilize yield (Lanari et al., 2014; Galvez et al., 2014; Bonada et al., 2018; Cole and Pagay, 2015). The synthesis and concentration of phenolic compounds, soluble solids, and anthocyanins were observed to be enhanced by water scarcity (Degaris et al., 2015; Ferrandino and Lovisolo, 2014; Conesa et al.,

2016). In contrast to rain fed vineyards Zarrouk et al. (2016) found lower levels of anthocyanins in the grape skins and less color in the wines. As a result of an increase in the berry's skin to pulp ratio. Bindon et al. (2011) claim that the benefits of water deficit in grapevines are directly linked to elements of wine quality such as color, flavour, and aroma. By promoting anthocyanin hydroxylation a water deficit can increase the accumulation of anthocyanins (Chaves et al., 2010). According to Romero et al. (2010), cv. Monastrell grapes exhibit a total grape phenolic compound concentration that is correlated with severe water stress. But according to a recent study Casassa et al. (2015), Cabernet Sauvignon grape varietals and wines with higher phenolic compound concentrations were produced when early and complete deficit irrigation was applied prior to veraison. Generally, red grapes with mild water stress have higher concentrations of these substances which enhances berry quality. Unfortunately these beneficial effects are reported to decrease once a specific level of water stress occurs (Romero et al., 2010). In a similar vein, Delgado Cuzmar et al. (2018) report that using less water can improve the chemical and sensory quality of wine. This is noteworthy given the possibility of declining water supplies due to climate change. According to Niculea et al. (2015), The concentration and content of phenolic compounds in response to extended deficit irrigation differ based on the type of berries that are developing and maturing. In comparison to the rain fed regime, the yield was significantly increased by the deficit irrigation (DI) and fully irrigated (FI) strategies, which improved vine water use by 18% and 27% respectively (Ramírez-Cuesta et al., 2023). Deficit irrigation (DI) is a technique that can assist modify the gap between technological and phenolic maturity in the context of the rain-fed technique, enhancing vine performance and production (Pérez-Álvarez et al., 2021).

# Plant Growth Regulators Affecting on Grapes Coloration:

### **Brassinosteroids (BRs)**

The sixth-largest phytohormone was found to be BRs, a sterol phytohormone, because of its beneficial effects on plant growth and stress resistance, even at extremely low concentrations (Sun et al., 2020). Vergara et al. (2018) state that exogenous applications of various BRs analogues to "Redglobe" grape clusters cause a notable change in the distribution of anthocyanin groups additionally to an increase in the color, soluble solids content, and total anthocyanins of the berries. Accordingly, Li et al. (2022) found that preharvest exogenous administration of 100 mmol. L<sup>-1</sup> jasmonic acid and 0.5 mg.L<sup>-1</sup> 2,4-epibrassinolide was an effective means of enhancing grape berry quality. The impact of exogenous BR applied to Cabernet Sauvignon clusters at a concentration of 0.4 mg/l on anthocyanin accumulation and anthocyanin biosynthesis gene expression (Xi et al., 2013 and Luan et al., 2013). Previous research found that pre-harvest administration of 2,4epibrassinolide at 0.4 and 0.6 mg.L<sup>-1</sup> significantly boosted proanthocyanidin accumulations and accelerated the metabolism of soluble sugars in the Cabernet Sauvignon and Merlot grapes, respectively (Xu et al., 2015; Yan et al., 2022).

### Jasmonic acid (JA)

Jasmonates (JAs) which are composed of methyl jasmonate (MeJA) a volatile derivative of jasmonic acid are thought to function as hormones in plants influencing several physiological functions, including growth, photosynthesis, reproductive development, and responses to biotic and abiotic stresses (Dar et al., 2015). According to reports, JA is crucial for causing stomatal closure, blocking Rubisco biosynthesis, impacting the absorption of N and P as well as the transportation of glucose and other organic compounds (El-kenawy, 2018; Gomi, 2020). Previous research has shown that MeJA treatments to vineyards increased the phenolic content, primarily anthocyanins, flavonols, and stilbenes on grape and wine, despite significant variations in the growing season and varieties (Gómez-Plaza et al., 2017; Portu et al., 2015, 2016, 2018). According to Flores et al. (2015), an evaluation of postharvest MeJA treatment revealed an increase in the 'Red Globe' cultivar's total phenolic and anthocyanin concentrations as well as antioxidant activity. Studies demonstrated the enhancement of anthocyanin and other polyphenolic compounds in grapevines and wines through the application

of MeJA treatments (Portu et al., 2015, 2016; Gil-Muñoz et al., 2017). M.E. García-Pastor et al. (2019) Demonstrate that MeJA 1mM treatments enhanced the total phenolic concentration at harvest by 1.3 and 1.5 times, respectively in 'Crimson' and 'Magenta' cultivars during 2016 experiment. Similarly, MeJA treatments were found to increase the concentration of phenolics during 2017 experiments in both cultivars at 0.1 mM was the most effective concentration. MeJA is currently applied directly to grapevines more frequently due to its positive effects on the synthesis of phenolic compounds in grapes (D'Onofrio et al., 2018). In particular, the anthocyanins, flavonols, and stilbenes in "Graciano" and "Garnacha" rose in the grapes after MeJA treatment (Portu et al., 2018; Portu et al., 2017). The amount of phenolic compounds in grapes after MeJA treatments differs mostly based on the response of the variety and the climate of the season (Portu et al., 2018; Gil-Muñoz et al., 2017).

### Salicylic acid (SA)

Ortho-hydroxyl benzoic acid, also known as salicylic acid (SA), is a phenolic naturally occurring plant growth regulator that is categorised as a growth promoter. According to Hayat et al. (2010), it has been discovered to be important in controlling plant growth, development, and vigour under both abiotic and biotic stresses. An effective agronomic method to produce table grapes with improved healthpromoting properties is by spraying grapevines with exogenous salicylic acid (Champa et al., 2015). Salicylic acid (SA) to postpone the ripening of "Superior Seedless" grape fruit clusters (Lo'ay, 2017). By reducing the activity of the enzymes that break down cell walls ascorbic acid at (6 mM) and salicylic acid at (4 mM) treatments prevented fruit cluster deterioration during shelf life while preserving phenolic compounds. According to Abdelaziz et al. (2022), the application of exogenous potassium silicate plus (SA) at 100 ppm showed to be the most successful treatment in maintaining the general quality of grapes that were stored. According to Champa et al. (2015), during cold storage the "Flame Seedless" cultivar's berry color, firmness, phenolic content, and organoleptic characteristics were all preserved by exogenous SA at 1.5 and 2 mM. Furthermore, SA foliar application at the pre-veraison stage of 'Sahebi' grapes increased the concentration of anthocyanins at harvest particularly malvidin-3glucoside the major anthocyanin in this cultivar as well as total phenolics and flavonoids (Oraei et al., 2019). Chen et al. (2006) found that after harvesting whole "Cabernet Sauvignon" berries in vivo infiltration of 150 \_M SA activated PAL by increasing PAL mRNA accumulation the synthesis of a new PAL protein and enzyme increased activity. The total phenolic concentration observed in berries from treated vines could also be attributed to the effects of salicylate treatment on PAL activity. Salicylate pre-harvest treatments would therefore increase antioxidant properties and health beneficial effects of table grape consumption, given the established role of phenolic compounds, including anthocyanins, in health beneficial properties (Xia et al., 2010; Flamini et al., 2013; Blanch et al., 2020). MeSa exogenous at 0.1 mM may increase bioactive compounds in table grapes that have antioxidant qualities which could improve the grapes' health benefits. Additionally, it could accelerate up the ripening process of grapevines and maintain qualitative characteristics throughout a long period of storage, according to (García-Pastor et al., 2020).

### Abscisic acid (ABA)

It is generally accepted that abscisic acid (ABA) naturally occurs in grape skins at the start of the ripening, when concentration of anthocyanins and other phenolic compounds also increases. It is in charge of anthocyanin production in grape berries (Boss et al., 1996a, b; Kobayashi et al., 2002). Although the UFGT gene is found in all grape varieties only red-colored cultivars express it (Boss et al., 1996b). Exogenous antagonist (S)-cis-abscisic acid (S-ABA) has been demonstrated in a number of applications to successfully increase the anthocyanin concentration in grape skin in previous studies. (Roberto et al., 2012, 2013; Koyama et al., 2014, 2019; Yamamoto et al., 2015; Domingues Neto et al., 2017). The use of S-ABA at the right concentration and time is essential for improving grape skin color development and these may differ based on the cultivar and application area (Peppi et al., 2006, 2007, 2008a).

By initiating the biosynthesis of anthocyanins, the application of S-ABA at different phases of grape maturation increases the amount of secondary metabolites in grapes, explaining the increase in anthocyanin (Tecchio et al., 2017). The application of S-ABA during pre- and veraison may have contributed to the rise in anthocyanin contents of 'Benitaka' table grapes. During the early stages of ripening, grape tissues are more susceptible to ABA stimulation for anthocyanin processing (Yamamoto et al., 2015). At the start of veraison ABA is crucial in controlling several genes including those connected to the signaling pathway and stimulation of anthocyanins (Gambetta et al., 2010). The "Isabella" grapes' anthocyanin contents and color were considerably enhanced by the foliar application of S-ABA at preveraison and post-veraison followed by a second application (Yamamoto et al., 2015). In table grapes, a single utilization of S-ABA results in a poorer grape color than multiple applications (Roberto et al., 2013).

Additionally, applying S-ABA at pre-veraison and veraison produced the best results. The accumulation of anthocyanin in the skin of grape berries has been suggested to be caused by sugars and ABA levels, which activate the gene expressions involved in anthocyanin production (Keller, 2015). The efficiency of the berries is increased when exogenous S-ABA is added around the time of veraison when the genes are already active. Prior research on "Crimson Seedless" revealed that the treated grapes' UFGT levels dropped three weeks after S-ABA was applied (Peppi et al., 2008b). Vanillylacetone increased the expression of genes that produced endogenous ABA which accelerated the biosynthesis of anthocyanins and caused The accumulation of grape berry anthocyanins in the skin (Enoki et al., 2017). According to Gagné et al. (2006), After véraison, endogenous ABA content rapidly increased and accelerated the ripening of

grape berries, including the production of anthocyanins (Pilati et al., 2017). Farag et al. (2018) suggested that using the composition containing lysophosphatidylethanolamine (LPE) commerical name (lisophos 70%) at 400 ppm plus magnesium nitrate at 1% (w/v) as well as Abscisic acid (ABA) comerical name (ProTone 10%) at 200 ppm to accelerate veraison and improve the "Crimson seedless" grapevine's berries quality under field conditions.

# Conclusion

Previous review observed that the environmental factors and plant growth regulators are more effective in influencing grapes coloration and also provide grape producers with feasible and applicable regime to produce high quality grape clusters.

### Acknowledgements

I am deeply grateful to assistant lecturer: Sara Reda Alaswad (Damanhour University) for her dedicated time, expertise and helping me to finish this review.

# **Conflicts of interest**

The author declares that he has no conflict of interest with respect to the publication of this article.

#### REFERENCES

- Abdelaziz, A. M., Shaarawi, S. A., & Ibrahim, W. M. (2022). Effect of pre-harvest application of salicylic acid, potassium silicate, and calcium chloride, on storability and quality attributes of table grape. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 50(4), 12940-12940. <u>https://doi.org/10.15835/nbha50412940</u>.
- Alston, J. M., & Sambucci, O. (2019). Grapes in the world economy. The grape genome, 1-24.
- Azuma, A. (2018). Genetic and environmental impacts on the biosynthesis of anthocyanins in grapes. *Horticulture Journal*, 87, 1–17. <u>https://doi.org/10.2503/hortj.OKD-IR02.</u>
- Azuma, A., Yakushiji, H., Koshita, Y., & Kobayashi, S. (2012). Flavonoid biosynthesis-related genes in grape skin are differentially regulated by temperature and light conditions. *Planta*. 236, 1067-1080. <u>https://doi.org/10.1007/s00425-012-1650-x</u>.
- Barnuud, N. N., Zerihun, A., Gibberd, M., & Bates, B. (2014). Berry composition and climate: responses and empirical models. *International Journal of Biometeorology*, 58, 1207–1223. <u>https://doi.org/10.1007/s00484-013-0715-2</u>,
- Bindon, K.A., Myburgh, P., Oberholster, A., Roux, K., & Du Toit, C.D. (2011). Response of grape and wine phenolic composition in *Vitis vinifera* L. cv. Merlot to variation in grapevine water status. *South African Journal of Enology and Viticulture*, 32(1), 71–88. <u>https://doi.org/10.21548/32-1-1368</u>.
- Blanch, G.P., Gómez-Jiménez, M.C., & del Castillo, M.L.R. (2020). Exogenous salicylic acid improves phenolic content and antioxidant activity in table grapes. *Plant Foods for Human Nutrition*, 75, 177–183. <u>https://doi.org/10.1007/s11130-019-00793-z.</u>
- Bonada, M., Buesa, I., Moran, M.A., & Sadras, V.O. (2018). Interactive effects of warming and water deficit on Shiraz vine transpiration in the Barossa Valley. *Australasian Open Access Journals*, 52, 117–133. <u>https://doi.org/10.20870/oeno-one.2018.52.2.1851</u>.
- Boss, P.K., & Davies, C. (2009). Molecular biology of anthocyanin accumulation in grape berries. In: Roubelakis-Angelakis, K.A. (Ed.), *Grapevine Molecular Physiology & Biotechnology*, 2<sup>nd</sup> ed. Springer, Berlin. 263–292. <u>https://doi.org/10.1007/978-90-481-2305-6\_10.</u>
- Boss, P.K., Davies, C., & Robinson, S. (1996a). Analysis of the expression of anthocyanin pathway genes in developing *Vitis vinifera* L. cv. Shiraz grape berries and the implications for pathway regulation. *Plant Physiology*, 111, 1059–1066. <u>https://doi.org/10.1104/pp.111.4.1059</u>.
- Boss, P.K., Davies, C., & Robinson, S. (1996b). Expression of anthocyanin biosynthesis pathway genes in red and white grapes. *Plant Molecular Biology*, 32, 565–569. <u>https://doi.org/10.1007/BF00019111</u>.
- Caravia, L.,Collins,C.,Petrie,P.R., & Tyerman,S.D. (2016). Application of shade treatments during Shiraz berry ripening to reduce the impact of high temperature. *Australian Journal of Grape Wine Research*, 22, 422–437. <u>https://doi.org/10.1111/ajgw.12248.</u>
- Casassa LF, Keller M., & Harbertson JF. (2015). Regulated deficit irrigation alters anthocyanins, tannins and sensory properties of cabernet sauvignon grapes and wines. *Molecules*, 20, 7820-7844. https://doi.org/10.3390/molecules20057820.
- Champa, W.A.H.; Gill, M.I.S.; Mahajan, B.V.C., & Arora, N.K. (2015). Preharvest salicylic acid treatments to improve quality and postharvest life of table grapes (*Vitis vinifera* L.) cv. Flame Seedless. *Journal of Food Science Technology*, 52, 3607–3616. <u>https://doi.org/10.1007/s13197-014-1422-7.</u>
- Chaves, M.M., Zarrouk, O., Francisco, R., Costa, J.M., Santos, T., Regalado, A.P., M.L. Rodrigues and C.M. Lopes (2010). Grapevine under deficit irrigation: hints from physiological and molecular data. *Annals of Botany*. 105(5), 661–676. <u>doi: 10.1093/aob/mcq030</u>
- Chen, J.Y., Wen, P.F., Kong, W.F., Pan, Q.H., Zhan, J.C., Li, J.M., Wan, S.B., & Huang, W.D. (2006). Effect of salicylic acid on phenylpropanoids and phenylalanine ammonia-lyase in harvested grape berries. *Postharvest Biol.ogy and Technology*, 40, 64–72. <u>https://doi.org/10.1016/j.postharvbio.2005.12.017</u>.
- Cheng, J. H., Wei, L. Z., & Wu, J. (2015). Effect of light quality selective plastic films on anthocyanin biosynthesis in vitis vinifera I. cv. yatomi rosa. *Journal of Agriculture, Science and Technology*, 17, 157–166. <u>http://jast.modares.ac.ir/article-23-57-en.html</u>.
- Chorti, E., Guidoni, S., Ferrandino, A., & Novello, V. (2010). Effect of different cluster sunlight exposure levels on ripening and anthocyanin accumulation in Nebbiolo grapes. *American Journal Enology and Viticulture*, 61, 23-30. <u>https://doi.org/10.5344/ajev.2010.61.1.23.</u>
- Cohen, S.D., & Kennedy, J. A. (2010). Plant metabolism and the environment: Implications for managing phenolics. *Critical Reviews in Food Science and Nutrition*, 50, 620– 643. <u>https://doi.org/10.1080/10408390802603441</u>.
- Cohen, S.D., Tarara, J.M., Gambetta, G.A., Matthews, M.A., & Kennedy, J.A. (2012). Impact of diurnal temperature variation on grape berry development, proanthocyanidin accumulation, and the expression of flavonoid pathway genes. *Journal of Experimental Botany*, 1–11. <u>https://doi.org/10.1093/jxb/err449</u>

- Cole, J., & Pagay, V. (2015). Usefulness of early morning stem water potential as a sensitive indicator of water status of deficit-irrigated grapevines (*Vitis vinifera* L.). Scientia Horticulturae, 191, 10–14. <u>https://doi.org/10.1016/j.scienta.2015.04.034</u>.
- Conde, C., Silva, P., Fontes, N., Dias, A.C.P., Tavares, R.M., Sousa, M.J., Agasse, A., Delrot, S., & Gerós, H. (2007). Biochemical changes throughout grape berry development and fruit and wine quality, *Global Science Books, Food*. 1,1–22.
- Conesa, M.R., Falagan, N., Jde, la Rosa, M., Aguayo, E., Domingo, R., & P´erez-Pastor, A. (2016). Post-veraison deficit irrigation regimes enhance berry coloration and health-promoting bioactive compounds in Crimson Seedless Table grapes. *Agricultural Water Management*, 163, 9–18. <u>https://doi.org/10.1016/j.agwat.2015.08.026</u>.
- Costantini, L., Malacarne, G., Lorenzi, S., Troggio, M., Mattivi, F., Moser, C., & Grando, M.S. (2015). New candidate genes for the fine regulation of the colour of grapes. *Journal of Experimental Botany*. 66, 4427 4440. <u>https://doi.org/10.1093/jxb/erv159.</u>
- D'Onofrio, C., Matarese, F., & Cuzzola, A. (2018). Effect of methyl jasmonate on the aroma of Sangiovese grapes and wines. *Food Chemistry*, 242, 352–361. <u>https://doi.org/10.1016/j.foodchem.2017.09.084.</u>
- Dar, T.A., Uddin, M., Khan, M.M.A., Hakeem, K.R., & Jaleel, H. (2015). Jasmonates counter plant stress: a review. *Environmental Experimental Botany*, 115, 49–57. <u>https://doi.org/10.1016/j.envexpbot.2015.02.010.</u>
- De Orduña, R.M. (2010). Climate change associated effects on grape and wine quality and production. *Food Research International*, 43, 1844-1855. <u>https://doi.org/10.1016/j.foodres.2010.05.001</u>.
- De Rosas, I., Ponce, M.T., Malovini, E., Deis, L., Cavagnaro, B., & Cavagnaro, P. (2017). Loss of anthocyanins and modification of the anthocyanin profiles in grape berries of Malbec and Bonarda grown under high temperature conditions. *Plant Science*, 258, 137-145. <u>https://doi.org/10.1016/j.plantsci.2017.01.015</u>.
- Degaris, K.A., Walker, R.R., Loveys, B.R., & Tyerman, S.D. (2015). Impact of deficit irrigation strategies in a saline environment on Shiraz yield physiology, water use, and tissue ion concentration. *Australian Journal of Grape and Wine Research*, 21, 468–478. <u>https://doi.org/10.1111/ajgw.12151</u>.
- Degu, A., Ayenew, B., Cramer, G. R., & Fait, A. (2016). Polyphenolic responses of grapevine berries to light, temperature, oxidative stress, abscisic acid and jasmonic acid show specific developmental-dependent degrees of metabolic resilience to perturbation. *Food Chemistry*, 212, 828–836. DOI: <u>10.1016/j.foodchem.2016.05.164.</u>
- Delgado Cuzmar, P., Salgado, E., Ribalta-Pizarro, C., Olaeta, J.A., López, E., Pastenes, C., & Cáceres-Mella, A. (2018). Phenolic composition and sensory characteristics of Cabernet Sauvignon wines: effect of water stress and harvest date. *International Journal of Food Science Technology*, 53, 1726-1735. <u>https://doi.org/10.1111/ijfs.13757</u>
- Demotes-Mainard, S., Péron, T., Corot, A., Bertheloot, J., Le Gourrierec, J., Pelleschi-Travier, S., Crespel, L., Morel, P., Huché-Thélier, L., Boumaza, R., Vian, A., Guérin, V., Leduc, N., & Sakr, S. (2016). Plant responses to red and far-red lights, applications in horticulture. *Environmental and Experimental Botany*, 121, 4–21. <u>http://dx.doi.org/10.1016/j.envexpbot.2015.05.010</u>
- Domingues Neto, F.J., Tecchio, M.A., Pimentel Junior, A., Vedoato, B.T.F., Lima, G.P., & Roberto, S.R. (2017). Effect of ABA on color of berries and in the anthocyanin accumulation and total phenolic compounds of 'Rubi' table grape (*Vitis vinifera*). *Australian Journal of Crop Science*, 1, 199–205. <u>https://doi.org/10.21475/ajcs.17.11.02.p269</u>,
- El-kenawy, M. A. (2018). Effect of spraying jasmonic acid and girdling on growth, yield and improving fruits quality of crimson seedless grapevine. *Egyptian Journal of Horticulture*, 45, 25–37. <u>https://doi.org/10.21608/EJOH.2018.2490.1041</u>
- Enoki, S., Hattori, T., Ishiai, S., Tanaka, S., Mikami, M., Arita, K., Nagasaka, S., & Suzuki, S. (2017). Vanillylacetone up-regulates expression of genes leading to anthocyanin accumulation by inducing endogenous abscisic acid in grape cell cultures. *Journal of Plant Physiology*, 219, 22-27. <u>https://doi.org/10.1016/j.jplph.2017.09.005</u>
- Faci, J.M., Blanco, O., Medina, E.T., & Martínez-Cob, A. (2014). Effect of post veraison regulated deficit irrigation in production and berry quality of Autumn Royal and Crimson table grape cultivars. Agricultural Water Management, 134, 73–83. <u>https://doi.org/10.1016/j.agwat.2013.11.009</u>
- Farag, K, M., Leila, A. Haggag., N. A. Abdel Ghany., N, M.N. Nagy., & Shehata. R.S. (2018). "Acceleration of Veraison and Enhancement of Berry Quality of "Crimson" Grapes By preharvest Treatments with Lysophosphatidylethanolamine, Protone and Magnesium". Assiut Journal of Agricultural Sciences. 49(2), 75-105. <u>https://doi.org/10.21608/AJAS.2018.11237</u>.
- Farag, K.M., Haikal, A.M., Nagy, N.M.N., & Shehata, R.S. (2011). Effect of modified Ethrel formulation and heat accumulation on berry coloration and quality of 'Crimson Seedless" grapes. A. Berry characteristics at harvest in relation to heat accumulation and number of pickings. *Journal of Agriculture and Environmental Sciences*, 10(3), 14-47. <u>https://doi.org/10.13140/RG.2.2.14795.26408</u>.

- Farag, K.M., Haikal, A.M., Nagy, N.M.N., & Shehata, R.S. (2012). Effect of modified Ethrel formulation and heat accumulation on berry coloration and quality of "Crimson Seedless" grapes. B. The interactions between treatments, type of heat accumulation, and number of pickings. *Journal of Agriculture and Environmental Sciences*, 11(3), 32-57.
- Ferrandino, A., & Lovisolo, C. (2014). Abiotic stress effects on grapevine (*Vitis vinifera* L.): focus on abscisic acidmediated consequences on secondary metabolism and berry quality. *Environmental and Expremental Botany*, 103, 138–147. <u>https://doi.org/10.1016/j.envexpbot.2013.10.012</u>.
- Flamini, Riccardo., Fulvio, Mattivi., Mirko, De Rosso., Panagiotis, Arapitsas., & Luigi, Bavaresco. (2013). "Advanced Knowledge of Three Important Classes of Grape Phenolics: Anthocyanins, Stilbenes and Flavonols" International Journal of Molecular Sciences, 14(10), 19651-19669. <u>https://doi.org/10.3390/ijms141019651</u>
- Flores, G., Blanch, G.P., & Ruiz del Castillo, M.L. (2015). Postharvest treatment with (-) and (+)-methyl jasmonate stimulates anthocyanin accumulation in grapes. *Food Science and Technology*, 62, 807–812. Doi: <u>10.1016/j.lwt.2014.12.033</u>.
- Fraga, H., Garcí ade Cortázar Atauri, I., & Santos, J.A. (2018). Viticultural irrigation demands under climate change scenarios in Portugal. *Agricultural Water Management*, 196, 66–74. <u>https://doi.org/10.1016/j.agwat.2017.10.023</u>.
- Fraga, H., Garcí ade Cortázar Atauri, I., Malheiro, A.C., & Santos, J.A. (2016). Modelling climate change impacts on viticultural yield, phenology and stress conditions in Europe. *Global Change Biology*, 22, 3774–3788. <u>https://doi.org/10.1111/gcb.13382</u>.
- Fraga, H., Malheiro, A.C., Moutinho-Pereira, J., & Santos, J. A. (2012). An overview of climate change impacts on European viticulture. *Food Energy Security*, 1, 94-110. <u>https://doi.org/10.1002/fes3.14</u>.
- Gagné , S ., Cluzet , S ., Merillon, J . M., & Geny, L. (2011). ABA initiates anthocyanin production in grape cell cultures. *Journal of Plant Growth Regulation*, 30, 1 -10. <u>https://doi.org/10.1007/s00344-010-9165-9</u>.
- Gagné, S., Estève, K., Deytieux, C., Saucier, C., & Gény, L. (2006). Influence of abscisic acid in triggering "véraison" in grape berry skins of *Vitis vinifera* L. cv. Cabernet Sauvignon. *Journal International Sciences Vigne Vin*, 40, 7-14. <u>https://doi.org/10.20870/oeno-one.2006.40.1.882.</u>
- Gaiotti, F., Pastore, C., Filippetti, I., Lovat, L., Belfiore, N., & Tomasi, D. (2018). Low night temperature at veraison enhances the accumulation of anthocyanins in Corvina grapes (*Vitis Vinifera* L.). Scientific Reports, 8 (1), 1– 13. doi: <u>https://doi.org/10.1038/s41598-018-26921-4.</u>
- Galvez, R., Callejas, R., Reginato, G., & Peppi, M.C. (2014). Irrigation schedule on table grapes by stem water potential and vapor pressure deficit allows to optimize water use. *Ciencia Tecnica Vitivinicola*, 29, 60–70. https://doi.org/10.1051/ctv/20142902060.
- Gambetta, G.A., Matthews, M.A., Shaghasi, T.H., Mcelrone, A.J., & Castellarin, S.D. (2010). Sugar and abscisic acid signaling orthologs are activated at the onset of ripening in grape. *Planta*, 232, 219–234. https://doi.org/10.1007/s00425-010-1165-2.
- Gao-Takai, M., A. Katayama-Ikegami., S. Nakano., K. Matsuda., & H. Motosugi. (2017). Vegetative growth and fruit quality of Ruby Roman' grapevines grafted on two species of rootstock and their tetraploids. *Horticulture Journal*, 86, 171–182. <u>https://doi.org/10.2503/hortj.OKD-009</u>
- García-Pastor, M. E., Serrano, M., Guillén, F., Castillo, S., Martínez-Romero, D., Valero, D., & Zapata, P. J. (2019). Methyl jasmonate effects on table grape ripening, vine yield, berry quality and bioactive compounds depend on applied concentration. *Scientia horticulturae*, 247, 380-389. <u>https://doi.org/10.1016/j.scienta.2018.12.043</u>
- García-Pastor, María E., Pedro, J. Zapata., Salvador, Castillo., Domingo, Martínez-Romero., Daniel, Valero., María, Serrano., & Fabián, Guillén. (2020). Preharvest Salicylate Treatments Enhance Antioxidant Compounds, Color and Crop Yield in Low Pigmented-Table Grape Cultivars and Preserve Quality Traits during Storage. Antioxidants 9. 9, 832. <u>https://doi.org/10.3390/antiox9090832</u>
- Gil-Muñoz, R., Fernández-Fernández, J.I., Crespo-Villegas, O., & Garde-Cerdán, T. (2017). Elicitors used as a tool to increase stilbenes in grapes and wines. *Food Research International*, 98, 34–39. <u>https://doi.org/10.1016/j.foodres.2016.11.035.</u>
- Gómez-Plaza, E., Bautista-Ortín, A.B., Ruiz-García, Y., Fernández-Fernández, J.I., & Gil- Muñoz, R. (2017). Effect of elicitors on the evolution of grape phenolic compounds during the ripening period. *Journal of the Science* of Food Agriculture, 97, 977–983.<u>https://doi.org/10.1002/jsfa.7823.</u>
- Gomi, K. (2020). "Jasmonic Acid: An Essential Plant Hormone" International Journal of Molecular Sciences 21. 4, 1261. <u>https://doi.org/10.3390/ijms21041261.</u>
- Greer, D.H., & Weedon, M.M. (2013). The impact of high temperatures on Vitis vinifera cv. Semillon grape vine performance and berry ripening. *Frontiers in Plant Science*, 4, 491. <u>https://doi.org/10.3389/fpls.2013.00491</u>

- Guan, L., Dai, Z., Wu, B.H., Wu, J., Merlin, I., Hilbert, G., Renaud, C., Gomès, E., Edwards, E., Li, S.H., & Delrot, S. (2016). Anthocyanin biosynthesis is differentially regulated by light in the skin and flesh of white-fleshed and teinturier grape berries. *Planta*, 243, 23-41. <u>https://doi.org/10.1007/s00425-015-2391-4</u>
- Guan, L., Li, J.H., Fan, P.G., Li, S.H., Fang, J.B., Dai, Z.W., Delrot, S., Wang, L.J., & Wu, B.H. (2014). Regulation of anthocyanin biosynthesis in tissues of a teinturier grape cultivar under sunlight exclusion. *American Journal* of Enology and Viticulture, 65(3), 363-374.<u>https://doi.org/10.5344/ajev.2014.14029</u>.
- Hayat, Q., Hayat, S., Irfan, M., & Ahmad, A. (2010). Effect of exogenous salicylic acid under changing environment: a review. Environmental and Experimental Botany, 68, 14–25. https://doi.org/10.1016/j.envexpbot.2009.08.005
- Jones, G. V., Reid, R., & Vilks, A. (2012). "Climate, grapes, and wine: Structure and suitability in a variable and changing climate," *in The Geography of Wine* (Dordrecht: Springer), 109–133. doi: <u>10.1007/978-94-007-0464-0 7</u>.
- Keller, M. (2015). The Science of Grapevines: Anatomy and Physiology, 2nd ed. Elsevier Academic Press, London.
- Kobayashi, S., Ishimaru, M., Hiraoka, C., & Honda, C. (2002). MYB-related genes of the Kyoho grape (Vitis labruscana) regulate anthocyanin biosynthesis. Plant Science, 215, 924–933. <u>https://doi.org/10.1007/s00425-002-0830-5</u>.
- Koyama, R., Assis, A.M., Colombo, R.C., Borges, W.F., Borges, R.S., Silvestre, J.P., Hussain, I., Shahab, M., Ahmed, S., Prudencio, S.H., Souza, R.T., & Roberto, S.R. (2019). Abscisic acid application affects the color and acceptance of the new hybrid 'BRS Melodia' seedless grape grown in a subtropical region. *HortScience*, 54 (6), 1055–1060. <u>https://doi.org/10.21273/HORTSCI13872-19</u>.
- Koyama, R., Assis, A.M., Yamamoto, L.Y., Borges, W.F., Borges, R.S., Prudencio, S.H., & Roberto, S.R. (2014). Exogenous abscisic acid increases the anthocyanin concentration of berry and juice from 'Isabel' grapes (*Vitis labrusca* L.). *HortScience*, 49 (4), 460–464. <u>https://doi.org/10.21273/HORTSCI.49.4.460</u>.
- Lee, J., & Skinkis, P. A. (2013). Oregon 'Pinot noir' grape anthocyanin enhancement by early leaf removal. *Food Chemistry*, 139, 893–901. <u>https://doi.org/10.1016/j.foodchem.2013.02.022</u>
- Li, J., Javed, H.U., Wu, Z., Wang, L., Han, J., Zhang, Y., Ma, C., Jiu, S., Zhang, C., & Wang, S. (2022). Improving berry quality and antioxidant ability in 'Ruidu Hongyu' grapevine through preharvest exogenous 2,4-epibrassinolide, jasmonic acid and their signaling inhibitors by regulating endogenous phytohormones. *Frontiers Plant Science*, 13:1035022. <u>https://doi.org/10.3389/fpls.2022.1035022</u>
- Lo'ay, A.A. (2017). Preharvest salicylic acid and delay ripening of 'superior seedless' grapes. *Egyptian Journal of Basic Applied Sciences*, 4, 227–230. <u>https://doi.org/10.1016/j.ejbas.2017.04.006</u>,
- Luan, L. Y., Zhang, Z. W., Xi, Z. M., Huo, S. S., & Ma, L. N. (2013). Brassinosteroids regulate anthocyanin biosynthesis in the ripening of grape berries. S. Afr. J. Enol. Vitic. 34, 196–203. DOI: <u>https://doi.org/10.21548/34-2-1094.</u>
- Ma, Z., Li, W., Mao, J., Li, W., Zuo, C., Zhao, X., Dawuda, M.M., Shi, X., & Chen, B. (2019). Synthesis of light-inducible and light-independent anthocyanins regulated by specific genes in grape 'Marselan' (V. vinifera L.) PeerJ 7:e6521 <u>https://doi.org/10.7717/peerj.6521.</u>
- Niculea, M., Martínez-Lapuente, L., Guadalupe, Z., Sánchez-Diaz. M., Ayestarán, B., & Antolín, MC. (2015). Characterization of phenolic composition of *Vitis vinifera* L. 'Tempranillo' and 'Graciano' subjected to deficit irrigation during berry development. *Vitis*, 54, 9-16. <u>https://doi.org/10.5073/vitis.2015.54.9-16</u>.
- Oraei, M., Panahirad, S., Zaare-Nahandi, F., & Gohari, G. (2019). Pre-veraison treatment of salicylic acid to enhance anthocyanin content of grape (*Vitis vinifera* L.) berries. *Journal of the Science Food and Agriculture*, 99, 5946–5952. <u>https://doi.org/10.1002/jsfa.9869.</u>
- Pastore, C., Dal Santo, S., Zenoni, S., Movahed, N., Allegro, G., & Tornielli, G. B. (2017). Whole plant temperature manipulation affects flavonoid metabolism and the transcriptome of grapevine berries. *Frontiers Plant Science*, 8, 929. <u>https://doi.org/10.3389/fpls.2017.00929.</u>
- Peppi, M.C., Fidelibus, M.W., & Dokoozlian, N. (2006). Abscisic acid application timing and concentration affect firmness, pigmentation and color of 'Flame Seedless' grapes. *HortScience*, 41, 1440–1445. DOI: <u>https://doi.org/10.21273/HORTSCI.41.6.1440</u>
- Peppi, M.C., Fidelibus, M.W., & Dokoozlian, N. (2007). Application timing and concentration of abscisic acid affect the quality of 'Redglobe' grapes. *The Journal of the Horticultural Science and Biotechnology*, 82 (2), 304– 310. <u>https://doi.org/10.1080/14620316.2007.11512233</u>.
- Peppi, M.C., Fidelibus, M.W., & Dokoozlian, N. (2008a). Timing and concentration of abscisic acid applications affect the quality of 'Crimson Seedless' grapes. *International Journal of Fruit Science*, 7 (4), 71–83. <u>https://doi.org/10.1080/15538360802003324</u>.
- Peppi, M.C., Walker, M.A., & Fidelibus, M.W. (2008b). Application of abscisic acid rapidly upregulated UFGT gene expression and improved color of grape berries. *Vitis*, 47 (1), 11–14. <u>https://doi.org/10.5073/vitis.2008.47.11-14</u>.

- Pérez-Álvarez, E., Intrigliolo Molina, D., Vivaldi, G., García Esparza, MJ., Lizama Abad, V., & Alvarez Cano, MI. (2021). Effects of the irrigation regimes on grapevine cv. Bobal in a Mediterranean climate: I. Water relations, vine performance and grape composition. *Agricultural Water Management*, 248, 1-13. <u>https://doi.org/10.1016/j.agwat.2021.106772.</u>
- Pilati, S., Bagagli, G., Sonego, P., Moretto, M., Brazzale, D., Castorina, G., Simoni, L., Tonelli, C., Guella, G., Engelen, K., Galbiati, M., & Moser, C. (2017). Abscisic Acid Is a Major Regulator of Grape Berry Ripening Onset: New Insights into ABA Signaling Network. *Frontiers Plant Science*, 8, 1093. <u>https://doi.org/10.3389/fpls.2017.01093.</u>
- Poni, S., Gatti, M., Palliotti, A., Dai, Z., Duch<sup>ene</sup>, E., Truong, T.-T., Ferrara, G., Matarrese, A.M.S., Gallotta, A., Bellincontro, A., Mencarelli, F., & Tombesi, S. (2018). Grapevine quality: a multiple choice issue. *Scientia Horticulturae*, 234, 445–462.<u>https://doi.org/10.1016/j.scienta.2017.12.035</u>.
- Portu, J., López, R., Baroja, E., Santamaría, P., & Garde-Cerdán, T. (2016). Improvement of grape and wine phenolic content by foliar application to grapevine of three different elicitors: methyl jasmonate, chitosan, and yeast extract. *Food Chemistry*, 201, 213–221. <u>https://doi.org/10.1016/j.foodchem.2016.01.086</u>.
- Portu, J., López, R., Ewald, P., Santamaría, P., Winterhalter, P., & Garde-Cerdán, T. (2018). Evaluation of Garnache, Graciano and Tempranillo grape stilbene content after field applications of elicitors and nitrogen compounds. *Journal of the Science of Food and Agriculture*, 98, 1856–1862. <u>https://doi.org/10.21548/34-2-1094.</u>
- Portu, J., López, R., Santamaría, P., & Garde-Cerdán, T. (2017). Elicitation with methyl jasmonate supported by precursor feeding with phenylalanine: effect on garnacha grape phenolic content. *Food Chemistry*, 237, 416–422.doi: <u>10.1016/j.foodchem.2017.05.126.</u>
- Portu, J., Santamaría, P., López-Alfaro, I., López, R., & Garde-Cerdán, T. (2015). Methyl jasmonate foliar application to tempranillo vineyard improved grape and wine phenolic content content. *Journal of Agricultural and Food Chemistry*, 63 2328.2337.DOI: 10.1021/jf5060672.
- Ramírez-Cuesta, J. M., Intrigliolo, D. S., Lorite, I. J., Moreno, M. A., Vanella, D., Ballesteros, R., Hernández-López, D., & Buesa, I. (2023). Determining grapevine water use under different sustainable agronomic practices using METRIC-UAV surface energy balance model.n*Agricultural Water Management*, 281, 108247. <u>https://doi.org/10.1016/j.agwat.2023.108247.</u>
- Roberto, S.R., Assis, A.M., Yamamoto, L.Y., Miotto, L.C.V., Sato, A.J., Koyama, R., & Genta,W. (2012). Application timing and concentration of abscisic acid improve color of Benitaka' table grape. *Scientia Horticulturae*, 142, 44–48. <u>https://doi.org/10.1016/j.scienta.2012.04.028</u>.
- Roberto, S.R., De Assis, A.M., Yamamoto, L.Y., Miotto, L.C.V., Koyama, R., Sato, A.J., & Borges, R.S. (2013). Ethephon use and application timing of abscisic acid for improving color of 'Rubi' table grape. *Pesquisa Agropecuaria Brasileira*, 48 (7), 797–800. <u>https://doi.org/10.1590/S0100-204X2013000700013</u>.
- Rocchetti, G., Ferrari, F., Trevisan, M., & Bavaresco, L. (2021). Impact of climatic conditions on the resveratrol concentration in blend of Vitis vinifera L. cvs. Barbera and Croatina grape wines. Molecules. 26, 401. <u>https://doi.org/10.3390/molecules26020401.</u>
- Rodrigues, M.L. (2010). Grapevine under deficit irrigation: Hints from physiological and molecular data. *Annals of Botany*, 105, 661-676. <u>https://doi.org/10.1093/aob/mcq030</u>.
- Romero, P., Fernández-Fernández, JI., & Martínez-Cutillas A. (2010). Physiological thresholds for efficient regulated deficit irrigation management in wine grapes grown under semiarid conditions. *American Journal of Enology and Viticulture*, 61, 300-312. <u>https://doi.org/10.5344/ajev.2010.61.3.300.</u>
- Sadras, V.O., & Petrie, P.R. (2011). Climate shifts in south-eastern Australia: Early maturity of Chardonnay, Shiraz and Cabernet Sauvignon is associated with early onset rather than faster ripening. *Australian Journal of Grape Wine Research*, 17, 199–205.<u>https://doi.org/10.1111/j.1755-0238.2011.00138.x.</u>
- Shinomiya, R., Fujishima, H., Muramoto, K., & Shiraishi, M. (2015). Impact of temperature and sunlight on the skin coloration of the 'Kyoho' table grape. Science Horticulture, 193, 77-83. <u>https://doi.org/10.1016/j.scienta.2015.06.042</u>.
- Stocker, T. F., Qin, D., Plattner, G.-K., Tignor, M. M. B., Allen, S. K., Boschung, J., Nauels, A., Xia, Y., Bex, V., & Midgley, P. M. (2014). "Climate change 2013: The physical science basis", in Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change (Cambridge: Cambridge University Press), 1535. <a href="https://doi.org/10.1017/CB09781107415324">https://doi.org/10.1017/CB09781107415324</a>.
- Sugiura, T., M. Shiraishi, S. Konno., & A. Sato. (2018). Prediction of skin coloration of grape berries from air temperature. *Horticulture Journal*, 87, 18–25. <u>https://doi.org/10.2503/hortj.OKD-061.</u>

- Sun, Yujun., Yunhan, He., Ali, Raza. Irfan., Xinmeng, Liu., Qiaoqiao, Yu., Qian Zhang., & Deguang Yang. (2020). "Exogenous Brassinolide Enhances the Growth and Cold Resistance of Maize (*Zea mays* L.) Seedlings under Chilling Stress" *Agronomy*, 10(4), 488. <u>https://doi.org/10.3390/agronomy10040488.</u>
- Tecchio, M.A., Neto, F.J.D., Junior, A.P., Da Silva, M.J.R., Roberto, S.R., & Smarsi, R.C. (2017). Improvement of color and increase in anthocyanin content of 'Niagara Rosada'grapes with application of abscisic acid. *African Journal of Biotechnology*, 16 (25), 1400–1403. <u>https://doi.org/10.5897/AJB2017.16073</u>.
- Vergara, A.E., Díaz, K., Carvajal, R., Espinoza, L., Alcalde, J.A., & Pérez-Donoso, A.G. (2018). Exogenous Applications of Brassinosteroids Improve Color of Red Table Grape (Vitis vinifera L. Cv. "Redglobe") Berries. *Frontiers Plant Science*, 9, 363. <u>https://doi.org/10.3389/fpls.2018.00363.</u>
- Xi, Z. M., Zhang, Z. W., Huo, S. S., Luan, L. Y., Gao, X., Ma, L. N., & Fang, Y. L. (2013). Regulating the secondary metabolism in grape berry using exogenous 24-epibrassinolide for enhanced phenolics content and antioxidant capacity. *Food Chemistry*, 141(3), 3056-3065 <u>https://doi.org/10.1016/j.foodchem.2013.05.137</u>
- Xia, E.Q., Deng, G.F., Guo, Y.J., & Li, H.B. (2010). Biological activities of polyphenols from grapes. *International Journal of Molecular Sciences*, 11, 622–646. <u>https://doi.org/10.3390/ijms11020622.</u>
- Xu, F., Xi, Z. M., Zhang, H., Zhang, C. J., & Zhang, Z. W. (2015). Brassinosteroids are involved in controlling sugar unloading in Vitis vinifera "Cabernet Sauvignon" berries during véraison. *Plant Physiology and Biochemistry*, 94, 197–208. <u>https://doi.org/10.1016/j.plaphy.2015.06.005</u>.
- Yamamoto, L.Y., Koyama, R., de Assis, A.M., Borges, W.F.S., de Oliveira, I.R., & Roberto, S.R. (2015). Color of berry and juice of 'Isabel' grape treated with abscisic acid in different ripening stages. *Pesquisa Agropecuaria Brasileira*, 50 (12), 1160–1167. <u>https://doi.org/10.1590/S0100-204X2015001200005</u>.
- Yan, H. E., Yanli, S. U. N., Fangfang, Z. H. A. O., & Hongjun, D. A. I. (2022). Effect of exogenous brassinolides treatment on sugar metabolism of merlot grape berries. *Acta Horticulturae Sinica*, 49, 117. <u>https://doi.org/10.16420/j.issn.0513-353x.2020-0834.</u>
- Zarrouk, O., Brunetti, C., Egipto, R., Pinheiro, C., Genebra, T., Gori, A., Lopes, C.M., Tattini, M., & Chaves, M.M. (2016). Grape ripening is regulated by deficit irrigation/ elevated temperatures according to cluster position in the canopy. *Frontiers Plant Science*, 15, 1640. <u>https://doi.org/10.3389/fpls.2016.01640</u>.